

Artificial synthesis of carbohydrates from electrochemically fixed carbon dioxide

Received: 3 May 2025

Accepted: 14 November 2025

Published online: 2 January 2026



Jing Li^{1,2,4}, Kedang Chen^{1,2,4}, Nathan E. Soland³, Jindou Yang^{1,2}, Yuanzuo Gao^{1,2}, Seonjeong Cheon^{1,2}, Yuming Su^{1,2}, Peidong Yang³✉ & Hailiang Wang^{1,2}✉

Sustainable synthesis of C₅₊ carbohydrates from CO₂ remains challenging due to the complexity of controlled CO₂ reduction and carbon–carbon coupling. Biochemical approaches can convert primary CO₂ reduction products into C₅₊ carbohydrates, but are often constrained by lengthy reaction periods, low production rates and system complexity. Here we present a two-step electrochemical reduction–formose reaction method that uses hydroxymethanesulfonate (HMS) as a more stable surrogate for formaldehyde to facilitate the direct synthesis of C₅₊ carbohydrates from electrochemically fixed CO₂. Using cobalt tetraaminophthalocyanine molecules supported on multiwalled carbon nanotubes as an electrocatalyst, we achieve an HMS Faradaic efficiency of ~12% at a total current density of 150 mA cm⁻². Employing direct CO reduction increases the Faradaic efficiency to ~25% with over 63% carbon efficiency. The produced HMS enables an efficient formose reaction under mild conditions reaching a yield of 20.4% for C₅₊ carbohydrates. The CO₂-derived HMS also demonstrates its versatility as a formaldehyde surrogate in other reactions for synthesizing various valuable chemical products, promising a new approach for feeding advanced chemical synthesis with electrochemically fixed CO₂ via the intercepted formaldehyde intermediate.

Converting CO₂ into fuels and feedstock chemicals using renewable energy offers a promising solution to mitigating greenhouse gas emissions and achieving a circular carbon economy^{1–4}. While substantial progress has been made in the development of electro- and photocatalytic systems that can produce various C₁ and C₂ products such as CO, formate, methanol, methane, ethylene and ethanol^{5–10}, new chemistry is still needed for generating more complex and valuable products. Long-chain (C₅₊) carbohydrates are an important target because they are the molecular building blocks of life and have a wide range of applications in the food, pharmaceutical and biotechnology industries^{11–14}. Upgrading CO₂ to C₅₊ carbohydrates via a single electro- or photocatalytic step, requiring extensive C–C coupling and delicate control in reduction degree, is extremely difficult^{15,16}. Consequently, the

reported synthesis of C₅₊ carbohydrates from CO₂ usually comprises at least two steps, beginning with either thermochemical or electrochemical reduction of CO₂ to C₁ or C₂ acids or alcohols, followed by multiple biochemical steps incorporating fermentation or enzyme catalysis to yield carbohydrates (Fig. 1a)^{17–19}. Thermochemical CO₂ hydrogenation as the first step is highly efficient but requires harsh high-temperature and high-pressure reaction conditions and an additional pre-step to generate H₂. The subsequent biochemical process is effective but falls short in production rate, yield and post-reaction separation, due to the reaction period (lasting a few days) and the multiple necessary reaction steps. Therefore, efficient and fast synthesis of C₅₊ carbohydrates from electrochemically fixed CO₂ through simple processes is highly desirable.

¹Department of Chemistry, Yale University, New Haven, CT, USA. ²Energy Sciences Institute, Yale University, West Haven, CT, USA. ³Department of Chemistry, University of California, Berkeley, CA, USA. ⁴These authors contributed equally: Jing Li, Kedang Chen. ✉e-mail: p_yang@berkeley.edu; hailiang.wang@yale.edu

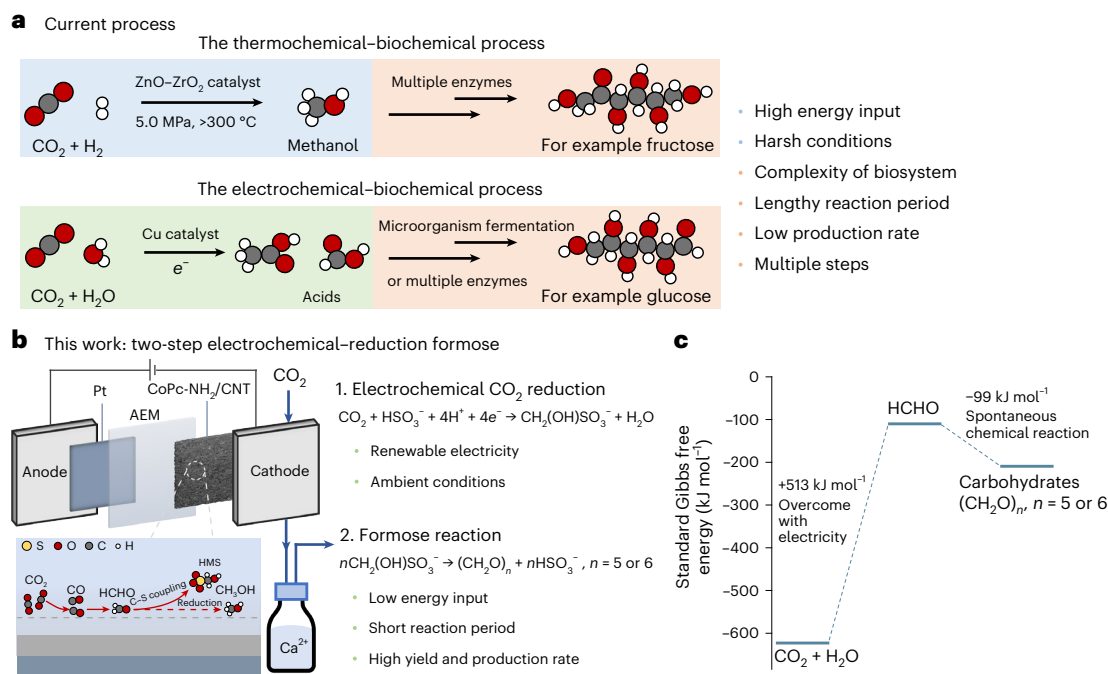


Fig. 1 | Comparing different pathways from CO_2 to carbohydrates.

a, Previously reported thermochemical-biochemical and electrochemical-biochemical processes. The shading represents different chemical steps: blue for thermochemical steps, green for electrochemical steps and orange for biochemical steps.

b, This work: a two-step electrochemical reduction-formose process. **c**, Free energy change for the two-step carbohydrate synthesis involving electrochemical CO_2 reduction followed by the formose reaction.

The formose reaction, which uses formaldehyde as the reactant, is the only known abiotic system capable of producing C_{5+} carbohydrates from a C_1 feedstock in a single step^{15,20,21}. This reaction can achieve high formaldehyde-to-carbohydrate conversion within a short duration of just a few hours and is suitable for large-scale production. From the perspective of energy consumption, it makes sense to couple the formose reaction with electrochemical reduction of CO_2 to formaldehyde for sustainable carbohydrate production (Fig. 1b), because the free energy cost of converting CO_2 into carbohydrates all falls in the CO_2 -to-formaldehyde step (Fig. 1c), which can be nicely overcome with renewable electricity²². However, effective formaldehyde production from CO_2 electroreduction has not yet been realized. This is likely to be because few electrocatalysts can reduce CO_2 by more than two electrons and formaldehyde is prone to further reduction or disproportionation under electrochemical conditions in aqueous media. Cobalt phthalocyanine (CoPc)-based materials are perhaps the only effective electrocatalysts for reducing CO_2 via formaldehyde; however, the reduction does not stop at formaldehyde due to the low thermodynamic and kinetic barriers for its further reduction to methanol^{23–25}. Although formaldehyde has been occasionally observed among the products of CoPc-catalysed CO_2 or CO electroreduction, its yield is very low^{26,27}. Stabilization of formaldehyde in electrochemical CO_2 reduction remains a major obstacle for its use in the synthesis of more valuable products.

In this Article, we develop a strategy that uses hydroxymethanesulfonate (HMS) as a more stable surrogate for formaldehyde to enable the direct synthesis of C_{5+} carbohydrates from electrochemically fixed CO_2 . Leveraging the known chemical reaction between formaldehyde and sulfite to form HMS, we manage to intercept the formaldehyde intermediate generated during CO_2 electroreduction catalysed by cobalt tetraaminophthalocyanine (CoPc- NH_2) molecules supported on multiwalled carbon nanotubes (CNTs). A Faradaic efficiency (FE) of ~12% is achieved for HMS at a total current density of 150 mA cm^{-2} , improving the state-of-the-art CO_2 -to-HMS conversion rate by one order of magnitude. The FE can be further improved to ~25% with an appreciable

carbon efficiency of over 60% when employing direct CO reduction. We then use an HMS solution generated from CO_2 electrolysis for the formose reaction, resulting in a yield of 20.4% for C_{5+} carbohydrates. We show that CO_2 -derived HMS can serve as a formaldehyde surrogate to perform the reactions of symmetric C–C coupling, asymmetric C–C coupling and C–N coupling for synthesizing various valuable chemicals such as ethylene glycol, sulfonated acetone formaldehyde (SAF) polycondensate and formaldoxime.

Results and discussion

Intercepting formaldehyde using sulfite

We first sought to verify the feasibility of the concept of intercepting formaldehyde with sulfite during CO_2 electrolysis catalysed by CoPc- NH_2 /CNT, which is one of the few electrocatalysts with stable activity towards reducing CO_2 to methanol^{24,28}. We employed our recently reported three-compartment flow electrolyser system with a gas diffusion electrode²⁹. This can achieve a high current density for CO_2 reduction to methanol, to ensure sufficient formaldehyde generation for capture. Galvanostatic electrolysis was performed in a CO_2 -saturated aqueous KHCO_3 electrolyte containing Na_2SO_3 at a moderate total current density of 150 mA cm^{-2} . In addition to typical products such as H_2 , CO and methanol, HMS was identified in the electrolyte by singlet peaks at -4.33 ppm in ^1H NMR and -74.25 ppm in ^{13}C NMR (Fig. 2a and Supplementary Fig. 1). This confirms successful interception of CO_2 -derived formaldehyde with sulfite via a known reaction involving nucleophilic attack of the sulfur in sulfite on the carbonyl of formaldehyde. We then varied the sulfite concentration (including bisulfite unless otherwise stated) from 0.2 to 0.8 M to examine its effect on HMS production. The measured FE of HMS increased from 4.6% at 0.2 M to 11.8% at 0.4 M (Supplementary Fig. 2). However, increasing the sulfite concentration further to 0.6 and 0.8 M reduced the formation rate and selectivity of both HMS and methanol while enhancing CO production. This is likely to be because the electrode potential became increasingly positive with higher electrolyte concentrations. To gain mechanistic information, we compared HMS and methanol

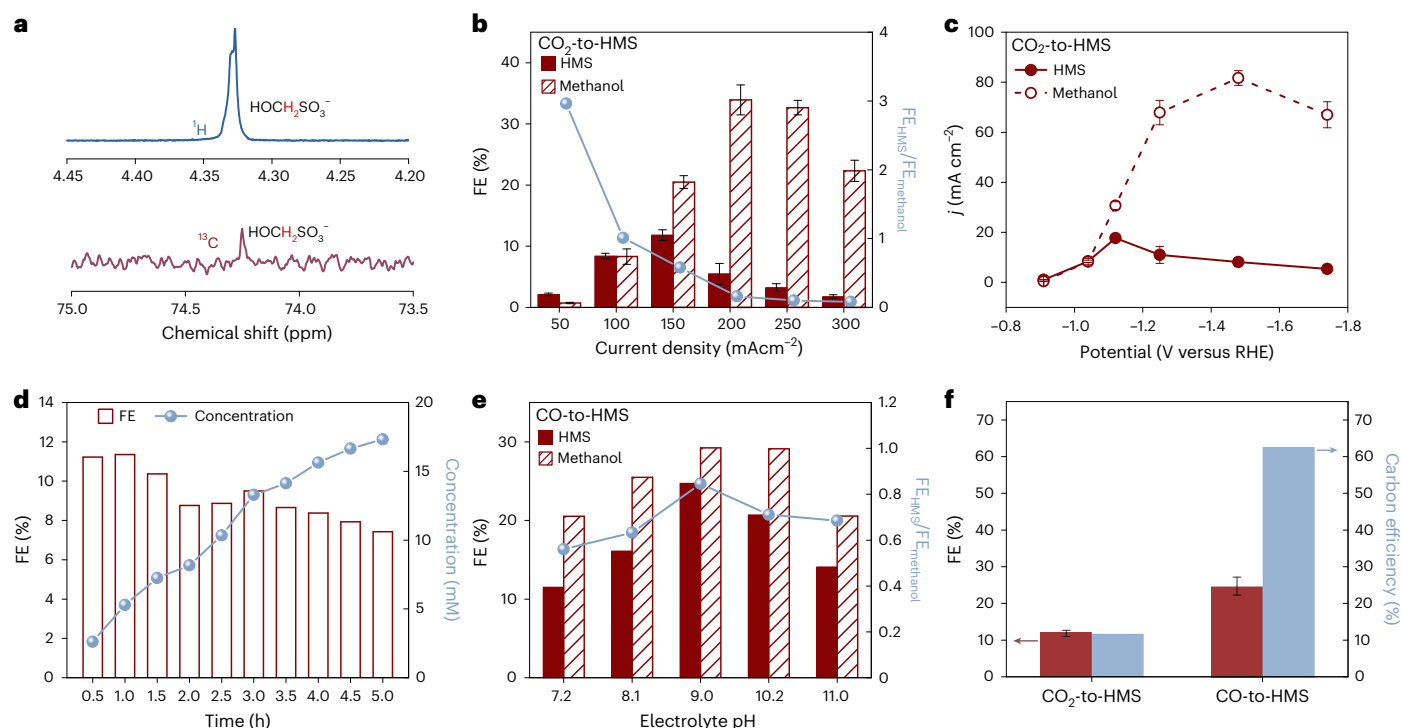


Fig. 2 | HMS production from CO₂ or CO electroreduction in the presence of sulfite. a, Detection of HMS by ¹H NMR and ¹³C NMR. **b**, FEs of HMS and methanol and their ratios for CO₂ reduction catalysed by 15 wt% CoPc-NH₂/CNT in an aqueous electrolyte containing 0.3 M KHCO₃ + 0.4 M Na₂SO₃ at various current densities. **c**, Partial current densities of HMS and methanol versus electrode potential for reactions in **b**. **d**, FE and concentration of HMS during 5 h CO₂ reduction electrolysis at 150 mA cm⁻². **e**, FEs of HMS and methanol and their

ratios versus electrolyte pH for CO reduction catalysed by 10 wt% CoPc-NH₂/CNT at -1.35 V versus SHE in an aqueous electrolyte containing 0.1 M KHCO₃ + 0.4 M Na₂SO₃ with its pH adjusted to different values. **f**, Comparison of FE and carbon efficiency between CO₂-to-HMS and CO-to-HMS processes, corresponding to the maximum HMS FE conditions in **b** and **e**, respectively. Data are presented as mean values and error bars represent standard deviations (*n* = 3 replicates).

formation rates in the electrolyte with and without sulfite under otherwise identical conditions (Supplementary Fig. 3). The methanol formation rate in the sulfite-containing electrolyte was found to be ~60% of that in the sulfite-free electrolyte, suggesting a partial interception of the methanol formation pathway. The total formation rate of methanol and HMS with sulfite was found to be comparable to the methanol formation rate without sulfite, indicating that both methanol and HMS are formed through a common formaldehyde intermediate. This result supports the understanding that HMS formation follows the CO₂-to-formaldehyde pathway.

Based on the optimized electrolyte condition, we conducted current-dependent electrolysis investigations. HMS began to form at 50 mA cm⁻² with an FE of 2.1%, accompanied by a small amount of methanol detected with an FE of 0.7% (Fig. 2b,c and Supplementary Fig. 4). The high formaldehyde interception efficiency of 82% is a clear indication that the C–S coupling reaction between sulfite and formaldehyde outcompetes the electroreduction of formaldehyde to methanol at this electrode potential. As the total current density was increased stepwise from 50 to 150 mA cm⁻², both FEs of HMS and methanol gradually increased, reaching the highest FE of 12% for HMS with a partial current density of 17.8 mA cm⁻² (Fig. 2b,c). This is nearly a 3-fold improvement in selectivity and a 13-fold increase in formation rate compared with a previous report using a Cu catalyst (4% FE and 1.3 mA cm⁻² for HMS production)³⁰. Further increasing the total current density resulted in a lower formation rate and selectivity for HMS production. Across the entire current density range explored, the total production rate of methanol and HMS combined follows a volcano shape, as commonly seen for CO₂ or CO electroreduction, whereas the HMS/methanol ratio monotonically decreased with increasing total current density (that is, increasingly negative electrode potential). This is due to the fact

that the C–S coupling reaction is a chemical step while its competing reaction, namely the reduction of formaldehyde to methanol, is electrochemically driven. At the optimal current density of 150 mA cm⁻², the catalyst was able to operate continuously for 5 h with a stable potential profile and an overall methanol FE of ~10%, affording a 17.5 mM HMS solution (Fig. 2d and Supplementary Fig. 5). The slight decrease of HMS FE over time can be attributed to consumption of HCO₃⁻ that is known to be a secondary proton source for the rate-determining *CO-to-*CHO step in formaldehyde formation²³.

Since HMS formation via formaldehyde–sulfite coupling is ultimately dependent on formaldehyde production, we sought to further improve the synthesis efficiency by conducting direct CO electrolysis, as our recent work reported a high CO-to-methanol FE of 66% catalysed by CoPc-NH₂/CNT (ref. 23). Conducting CO reduction in an H-cell containing 0.1 M aqueous KHCO₃ with 0.4 M Na₂SO₃ (pH = 8.6), we observed an FE of 20% for HMS at the optimal potential of -0.83 V versus the reversible hydrogen electrode (RHE) (Supplementary Fig. 6). We then varied the electrolyte pH from 7.2 to 11.0 using KOH or H₂SO₄. Measured at the potential of -1.35 V versus the standard hydrogen electrode (SHE), HMS and methanol FEs as well as the ratio of FE_{HMS}/FE_{methanol} exhibited a volcano-shaped trend across the pH range (Fig. 2e and Supplementary Fig. 7). This is a result of the interplay between several factors: (1) the competition between CO reduction and H₂ evolution is affected by both pH and HCO₃⁻ concentration²³, (2) pH controls the partition between sulfite and bisulfite, which may affect the C–S coupling reaction rate, (3) formaldehyde reduction to methanol, the competing reaction to HMS formation, is affected by pH and (4) higher pH (>9) favours the dissociation of HMS. At the optimal pH of 9.0, we achieved an FE of 25% and a notable overall carbon efficiency of 63% for HMS production, with great improvement over direct CO₂

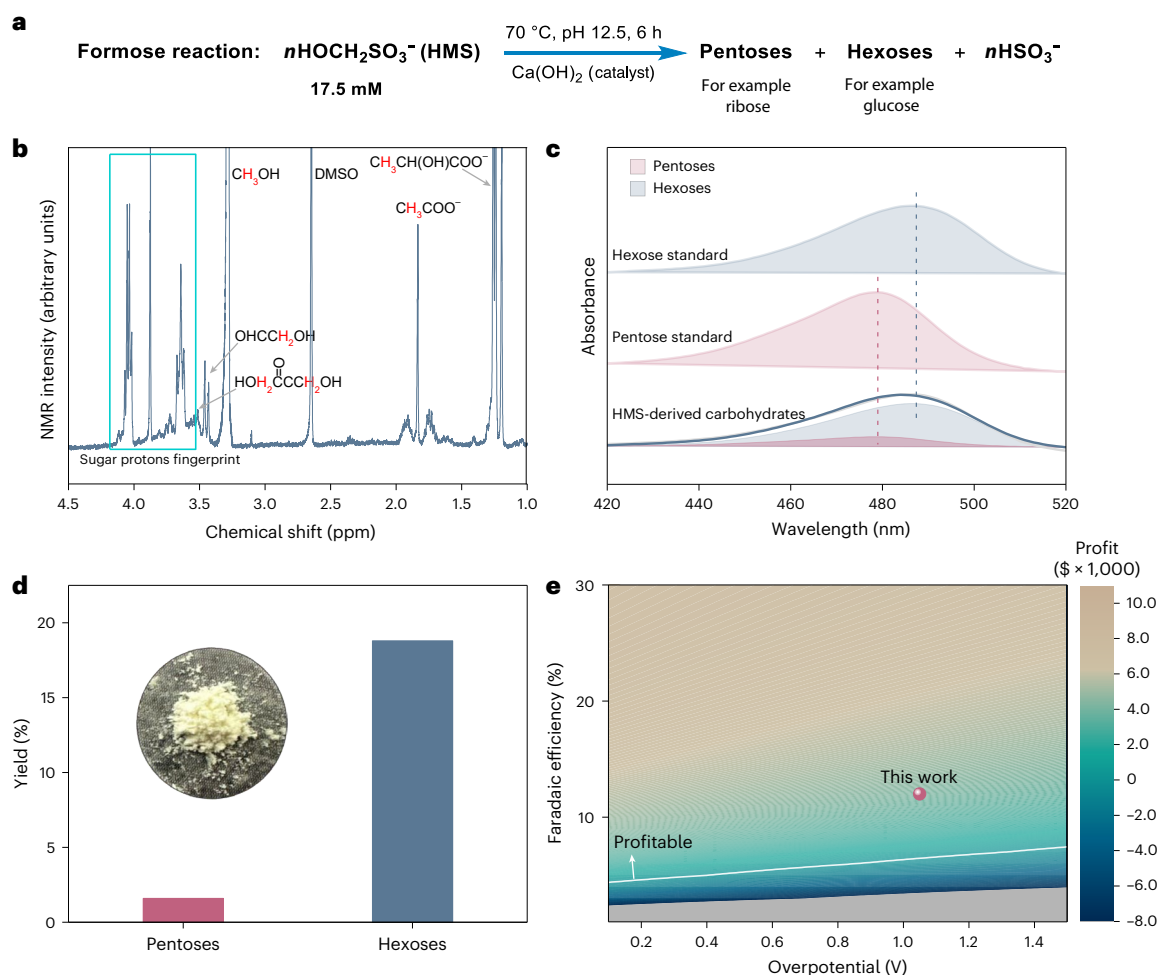


Fig. 3 | Carbohydrate synthesis via the formose reaction of a 17.5 mM HMS solution from CO₂ electrolysis. **a**, Scheme of the formose reaction using HMS as the reactant. **b**, ¹H NMR analysis of products from the formose reaction. **c**, Ultraviolet–visible absorption analysis of products from the formose reaction compared with hexose and pentose standards using the phenol–sulfuric acid

method. **d**, Yield of pentoses and hexoses from the formose reaction. The inset shows the separated carbohydrate solid. **e**, Techno-economic assessment of the profit of producing 1 t of C₅₊ carbohydrates from CO₂ as a function of the overpotential and Faradaic efficiency for HMS formation.

reduction (Fig. 2f). Since CO₂ can be electrochemically reduced to CO with near-unity FE^{9,31}, a tandem process to produce HMS from CO₂ via CO is of potential.

Coupling electrochemical CO₂ reduction with the formose reaction

With the success in capturing the unstable formaldehyde intermediate from CO₂ electroreduction catalysed by CoPc-NH₂/CNT in the form of HMS, we proceeded to the synthesis of carbohydrates utilizing the formose reaction. While numerous studies have reported that the formose reaction can convert formaldehyde into a spectrum of C₂ to C₅₊ carbohydrates under mild heating and alkaline conditions^{32–34}, a similar reaction using HMS as the starting reactant has not yet been reported. We first investigated the stability of HMS against pH at room temperature and found that HMS (50 mM, aqueous) is stable at pH of 8.8 or lower (Supplementary Fig. 8) with negligible decomposition into formaldehyde. This stability allows us to intercept the formaldehyde intermediate in the form of HMS under electrochemical conditions, preventing its further reduction to methanol. At a high pH of 12.5, over 99% of HMS decomposed within 6 h to release formaldehyde (Supplementary Fig. 8). This result clearly shows the feasibility of using HMS as an effective formaldehyde surrogate for the formose reaction in alkaline solutions. We then employed the aforementioned

CO₂ electrolysis solution containing 17.5 mM HMS for the formose reaction. Prior to reaction, an excess of 10 mM Ca²⁺ was added as the catalyst to the solution after precipitating CO₃²⁻ and SO₃²⁻. To expedite the formose reaction, glycolaldehyde was also added as the initiator to reach a concentration of 10 μM. The reaction was conducted at a mild temperature of 70 °C and a pH of 12.5 for 6 h (Fig. 3a).

We first performed ¹H NMR spectroscopy to analyse the reaction products. As shown in Fig. 3b and Supplementary Fig. 9, HMS was fully converted, with ~20% of it disproportioning to formate and methanol. We also observed lactic acid (27.2% yield) and acetic acid (1.4% yield), which are formed from retro-aldol breakdown of carbohydrates^{21,35}, as well as glycolaldehyde (0.8% yield) and dihydroxyacetone (3.2% yield) (Supplementary Fig. 10), which are important intermediates leading to the formation of C₅₊ carbohydrates³². The NMR peaks in the 3.5 to 4.2 ppm region correspond to the C-bonded protons in carbohydrate-like species and their derivatives (for example sugar acids). To quantify these higher-order carbohydrate products, we employed the phenol–sulfuric acid assay, a well-established colorimetric method for determining total C₅₊ monosaccharides³⁶. The total yield of C₅₊ carbohydrates was determined to be 20.4%, comprising 18.8% hexoses and 1.6% pentoses (Fig. 3b). Identification and quantification of each specific carbohydrate in this complex mixture remains a big challenge in the field^{21,37}. This is due to the lack of standard analytical techniques to

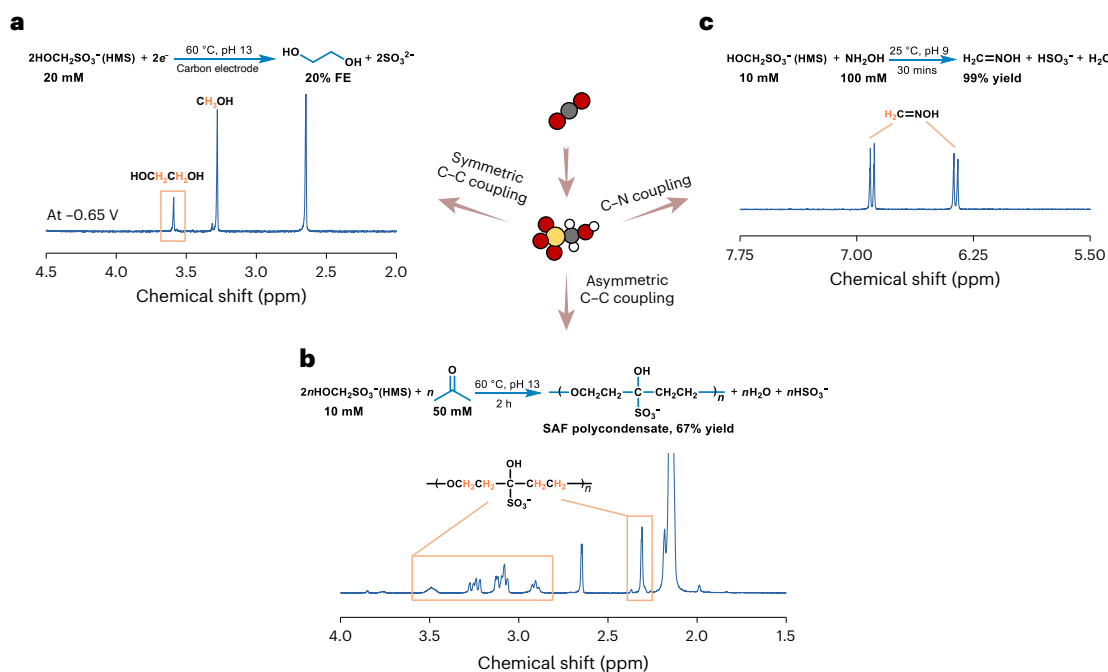


Fig. 4 | Extended application of HMS in various chemical syntheses. a, Electrochemical synthesis of ethylene glycol via symmetric C–C coupling. **b,** Chemical synthesis of sulfonated acetone formaldehyde polycondensate via asymmetric C–C coupling. **c,** Chemical synthesis of formaldoxime via C–N coupling.

effectively separate carbohydrate isomers and factors like the dynamic Lobry de Bruyn–Van Ekenstein transformation in formose reaction products that facilitates structural interconversion via aldose–ketose isomerization or epimerization³⁸.

To separate the carbohydrate products from the reaction medium, we acidified the post-reaction solution using HCl, evaporated all the liquid and used ethanol to redissolve the products (Supplementary Fig. 11). After evaporating the ethanol solvent, a crude carbohydrate powder was obtained with a yield of 45% (Fig. 3d inset). This value is roughly consistent with the yield estimated from the carbohydrate fingerprint region in the NMR spectrum but substantially exceeds the yield determined by the phenol–sulfuric acid assay. This is likely to be due to the presence of unseparated by-products, such as sugar acids and C₄ or branched carbohydrates, which have relatively high boiling points and similar physicochemical properties to the target carbohydrates. The sugar acids are generated primarily from the Cannizzaro reaction and degradation of carbohydrates formed during the formose reaction³⁵, whereas the branched carbohydrates form in the formose reaction through carbon–carbon bond formation at secondary carbon atoms³⁹. These by-products give NMR signals in the fingerprint region but are not detectable by the colorimetric method as they cannot form the requisite furfural derivatives. Based on the conservative carbohydrate yield of 20.4%, the overall energy efficiency of the CO₂-to-C₅₊ carbohydrates process is calculated to be approximately 0.8% (Methods), which is higher than that of the previously reported electrochemical–biochemical CO₂-to-glucose process (0.65%)¹⁸. Although this is lower than that of the recently reported CO₂-to-sorbitol conversion via a cascade of five enzyme-catalysed steps (3.5%)¹⁷, our electrochemical reduction–formose route still holds promise because it avoids delicate biosystems, delivers considerably higher reaction rates (Supplementary Table 1) and retains the possibility for further improvement in both HMS production from CO₂ and formose sugar production. However, we should note that the carbohydrates obtained in this work are not food or feed grade, and are likely to require downstream purification to comply with food-grade specifications. Our techno-economic analysis (TEA) demonstrates economic profitability for this process when capital,

operating and materials expenses are considered (Supplementary Note 1 and Supplementary Tables 2–4) (Fig. 3e). Our cradle-to-gate life cycle assessment (LCA) estimates that the global warming potential of this process is lower than that reported for conventional agriculture (Supplementary Fig. 12 and Supplementary Note 2)¹⁶, reflecting the possibility of reducing environmental impact. Combined TEA and LCA results highlight the plausibility of the electrochemical reduction–formose route as a sustainable approach for producing valuable C₅₊ carbohydrates.

It should be mentioned that Na₂SO₃ only functions as a mediator for the capture-and-release of formaldehyde rather than a co-reactant in carbohydrate formation. It captures formaldehyde as HMS during CO₂ electrolysis, and ends up as CaSO₃ precipitate prior to the formose reaction. The formose reaction thus consumes only the formaldehyde released from HMS and sulfite does not enter the carbohydrate product stream. No additional cost is imposed on downstream separations. If desired, cost can be further reduced by recycling sulfite. A simple recycling loop can be applied to recover Na₂SO₃ from CaSO₃ via acidification or thermal treatment to release SO₂ followed by chemical absorption (Supplementary Fig. 13). Additionally, the close match between the ¹H NMR spectra of carbohydrates derived from HMS and from commercial formaldehyde suggests that sulfite present as CaSO₃ does not participate in or alter the formose chemistry (Supplementary Fig. 14). Therefore, the use of Na₂SO₃ does not impact the economic feasibility of carbohydrate production or interfere with the formose reaction.

Extended applications of HMS in chemical synthesis

Given the success of using HMS for carbohydrate synthesis via the formose reaction, we sought to expand the scope of utilizing HMS as a formaldehyde surrogate in chemical synthesis. All reactions explored in this section were carried out with HMS concentrations of 20 mM or lower, which are readily achievable from our electrochemical CO₂ reduction reactions. We began with electroreductive synthesis of ethylene glycol, which is widely used as an antifreeze agent and a building block in the production of plastics⁴⁰. An early study demonstrated that

graphitic carbon can catalyse the electroreduction of formaldehyde to ethylene glycol in aqueous media⁴¹. We followed a recent study and employed a commercial carbon-black electrocatalyst in 0.1 M aqueous KOH at 60 °C in an H-cell to reduce HMS⁴², achieving a maximum FE of ~20% at a partial current density of 2.2 mA cm⁻² for ethylene glycol production at -0.65 V versus RHE (Fig. 4a and Supplementary Fig. 15). We observed that the reduction of formaldehyde to methanol dominated with an FE of 74%. This is likely to be due to the low concentration of HMS favouring single-molecule reduction over reductive dimerization of formaldehyde. Although the selectivity and formation rate are too low for practical application, this result successfully demonstrates the feasibility of using HMS to produce ethylene glycol via symmetric C-C coupling and provides an example of ethylene glycol electrosynthesis from CO₂.

Beyond electrochemical C-C coupling, we also explored chemical reactions between HMS and other reagents. As a surrogate for electrophilic formaldehyde, HMS is expected to be susceptible to nucleophilic attack, and so applicable to methylation and hydroxymethylation reactions. We first demonstrated the asymmetric C-C coupling reaction between HMS and an enolizable aldehyde or ketone. For example, reacting HMS with acetone in aqueous KOH at 60 °C produced a sulfonated acetone formaldehyde polycondensate with a yield of 67% (Fig. 4b). This product has widespread applications in the construction and materials industries as a superplasticizer, water reducer or dispersant⁴³. Additionally, we explored C-heteroatom coupling reactions for the use of HMS. As an example, formaldoxime was synthesized with a high yield of 99% by reacting HMS with hydroxylamine at room temperature in aqueous KHCO₃ to form C-N bonds (Fig. 4c). Formaldoxime is widely used as an intermediate in organic/pharmaceutical synthesis and as a ligand in analytical chemistry⁴⁴. These reaction examples collectively demonstrate the potential of using HMS as a reactive yet stable form of electrochemically fixed CO₂ for chemical synthesis.

Conclusions

In summary, we have demonstrated that formaldehyde captured in the form of HMS can enable a two-step electrochemical reduction-formose process for synthesizing valuable carbohydrates, such as pentoses and hexoses, using CO₂ as the carbon source. The successful extension of HMS as a formaldehyde surrogate to symmetric C-C, asymmetric C-C and C-N coupling reactions suggests a promising approach to bridging CO₂ electrochemical reduction with more advanced chemical and electrochemical reactions through capturing and stabilizing its active intermediates. Future work will focus on enhancing the rate and selectivity of HMS production from CO₂, optimizing carbohydrate selectivity in the HMS-based formose reaction and improving the separation of post-reaction products.

Methods

Materials

Materials used in this work were directly purchased from the following suppliers unless otherwise specified. Potassium hydroxide (99.98%) and acetone (>99.5%) were purchased from Fisher Chemical. Sodium sulfite (>98.0%), potassium bicarbonate (99%) and *N,N*-dimethylformamide (DMF, 99.8%) were purchased from Alfa Aesar. Calcium chloride (96%), hydroxylamine hydrochloride (99+%), D-glucose (ACS reagent) and D-galactose (99+%) were purchased from Acros Organics. Glycolaldehyde (crystalline) and Nafion solution (5 wt%) were purchased from Sigma Aldrich. 1,3-Dihydroxyacetone (97%), D-ribose (98%), D-fructose (99%), D-xylose (99+%) and D-mannose (99+%) were purchased from Thermo Fisher Scientific. Sodium hydroxymethanesulfonate (>97.0%) and D-arabinose (>99.0%) were purchased from Tokyo Chemical Industry. Carbon monoxide (99.3%), carbon dioxide (99.99%), argon (99.999%) and nitrogen (99.999%) gases were purchased from Airgas. Multiwalled CNTs were purchased from C-Nano (product number FT9100). The carbon paper support (Freudenberg, catalogue no.

H23C6) with microporous layer coating was purchased from the Fuel Cell Store. The electrolyte solutions were prepared using Milli-Q water (18.2 MΩ cm at 25 °C).

Preparation of CoPc-NH₂/CNT electrode

Details of the synthesis of CoPc-NH₂, purification of CNTs and preparation of CoPc-NH₂/CNT can be found in our previous work²³. Briefly, CoPc-NH₂ was synthesized in a solid-state reaction from 4-nitrophthalonitrile, CoCl₂·6H₂O and urea, catalysed by (NH₄)₆Mo₇O₂₄·4H₂O. CNTs were purified by calcination and HCl treatment. CoPc-NH₂ molecules were assembled on CNTs from their DMF solutions with the assistance of sonication and stirring. CoPc-NH₂/CNT electrodes with optimized molecular loadings of 15 wt% and 10 wt% were used for CO₂ and CO reduction, respectively^{23,29}. Optimized mass loadings of 1.2 and 0.4 mg cm⁻² for CoPc-NH₂/CNT electrodes on carbon paper were used for measurements in the flow cell and H-cell, respectively^{24,29}. To prepare CoPc-NH₂/CNT electrodes, 5 mg of CoPc-NH₂/CNT was mixed with 15 μl Nafion solution and 5 ml ethanol and this was followed by 30 min of sonication. The resulting ink was drop-casted onto a carbon paper support to achieve the desired catalyst loading, covering an area of approximately 1.0 cm². The electrode was then thoroughly dried for subsequent measurements using an infrared lamp.

Preparation of carbon catalyst electrode

The carbon catalyst electrode for ethylene glycol synthesis was prepared according to a reported method⁴². To prepare the carbon catalyst, 100 mg carbon black (Vulcan XC-72 R, Fuel Cell Store) was first purified using 100 ml of 5 wt% HCl under sonication for 30 min. The Vulcan carbon was then collected by filtration and washed with Milli-Q water until the supernatant reached pH 7. After freeze drying, the purified Vulcan carbon was annealed at 500 °C for 3 h under a 5:95 H₂/Ar atmosphere to eliminate oxygen-containing functional groups. A total of 10 mg of the pre-treated Vulcan carbon was mixed with 30 μl Nafion solution and 10 ml ethanol. This was followed by 30 min of sonication to create an ink, which was drop-casted onto carbon paper to achieve a mass loading of 0.5 mg cm⁻².

Electrochemical reactions

CO₂ electrolysis was performed in a three-channel flow electrolyser (Gaoss Union) with channel dimensions of 2 cm × 0.5 cm × 0.5 cm. The cathode and anode channels were separated by a piece of anion-conducting membrane (Selecion DSV, AGC). A platinum foil was used as the counter electrode and Ag/AgCl (0.199 V versus SHE, 4.0 M KCl, Pine Research Instrumentation) was used as the reference electrode. The electrolyte was 0.3 M CO₂-saturated aqueous KHCO₃ containing Na₂SO₃ with various concentrations. CO₂ gas was delivered into the cathode gas channel at a flow rate of 10.0 cm³ min⁻¹ using a mass flow controller (Alicat Scientific). The catholyte and anolyte flow rates were set to be 1.2 and 50 ml min⁻¹, respectively, using a peristaltic pump. CO electrolysis was performed in a home-made two-compartment H-cell separated by an anion-conducting membrane. A graphite rod (Sigma Aldrich, 99.999%) and Ag/AgCl were used as the counter electrode and reference electrode, respectively. The electrolyte was 0.1 M aqueous KHCO₃ containing 0.4 M Na₂SO₃. Prior to electrolysis, the catholyte was purged for 5 min by delivering CO gas at a flow rate of 50.0 cm³ min⁻¹. Then, CO gas was continuously supplied to the cathode compartment at a flow rate of 20.0 cm³ min⁻¹ for CO electrolysis. The symmetric C-C coupling reaction for ethylene glycol synthesis was carried out in the same H-cell. The electrolyte was 0.1 M aqueous KOH containing 20 mM HMS and the reaction was performed under Ar at 60 °C. All electrochemical experiments were performed using a Bio-Logic VMP3 Multichannel Potentiostat. Chronopotentiometry and chronoamperometry were used for the measurements in the flow cell and H-cell, respectively. The resistance between the working and

reference electrodes was determined by potentiostatic electrochemical impedance spectroscopy and was automatically compensated for during measurements.

Thermochemical reactions

Thermochemical reactions including the formose reactions, asymmetric C–C coupling and C–N coupling were conducted under controlled temperature and stirring conditions. For the formose reaction, the electrolyte containing 17.5 mM HMS after CO₂ electrolysis was used directly. Prior to reaction, CaCl₂ powder in an equimolar amount to SO₃²⁻ + CO₃²⁻ was gradually added to the electrolyte solution to precipitate CaSO₃ and CaCO₃. The pH of the solution was adjusted to 12.5 by adding 1.0 M aqueous KOH. Additional CaCl₂ and glycolaldehyde were then added to the solution to reach concentrations of 10 mM and 10 μM, respectively. The reaction was maintained at 70 °C for 6 h. Product separation, wherever applicable, was performed following the procedure illustrated in Supplementary Fig. 11. To prepare the aqueous formaldehyde solution for the formose reaction, paraformaldehyde was dissolved and hydrolysed in the Milli-Q water at 60 °C for 1 h. For the asymmetric C–C coupling reaction to synthesize sulfonated acetone formaldehyde polycondensate, commercial HMS and acetone were added to 20 ml 0.1 M aqueous KOH to reach final concentrations of 10 and 50 mM, respectively. This reaction was maintained at 60 °C for 2 h. For the C–N coupling reaction to synthesize formaldoxime, commercial HMS and hydroxylamine hydrochloride were added to 20 ml 0.1 M aqueous KHCO₃ to reach concentrations of 10 mM and 100 mM, respectively. This reaction was conducted at room temperature for 30 min.

Product quantification

Gas-phase products were analysed using a gas chromatograph (SRI 8610C) equipped with a flame ionization detector for CO quantification and a thermal conductivity detector for H₂ quantification. Liquid products were quantified using a Bruker AVIII 400 MHz NMR spectrometer with water suppression. NMR samples were prepared by mixing 450 μl of the resulting solution of each reaction with 50 μl of 10 mM dimethyl sulfoxide (Alfa Aesar, ≥99.9%) in D₂O (Sigma Aldrich, 99.9%) as the internal standard. The C₅₊ formose reaction products were quantified using a phenol–sulfuric acid assay. Calibration curves were obtained using aqueous solutions of 12–60 mg l⁻¹ glucose and 10–50 mg l⁻¹ ribose as representative standards for hexoses and pentoses, respectively (Supplementary Fig. 16). Typically, 2 ml of a carbohydrate standard or 4-fold diluted formose reaction sample was mixed with 50 μl of 80 wt% phenol aqueous solution, immediately followed by the addition of 5 ml concentrated sulfuric acid to produce furfural derivatives for colorimetric assay. Absorption spectra were recorded at 479 and 487 nm for pentoses and hexoses, respectively, using a Shimadzu UV-2600 ultraviolet–visible spectrophotometer.

Calculation of energy efficiency

For the CO₂-to-HMS process, the energy efficiency (EE_{electrochemical}) can be calculated as follows

$$EE_{\text{electrochemical}} = \frac{1.23 + (-E_0)}{E_{\text{applied}}} \times FE = \frac{1.23 + 0.07}{3.24} \times 12\% = 4.8\%$$

where E_{applied} is the cell voltage applied in the electrolysis, FE is the Faradaic efficiency of HMS and $E_0 = -0.07 V_{\text{RHE}}$ is the equilibrium potential for CO₂-to-formaldehyde conversion.

To determine the energy efficiency for the HMS-to-carbohydrates process in the formose reaction, we compare the enthalpy of combustion of the resulting carbohydrates to that of formaldehyde. As carbohydrates containing the same number of carbon atoms have nearly identical enthalpies of combustion, ribose and glucose were selected as representatives. According to data from the National Institute

of Standards and Technology and the CRC Handbook of Chemistry and Physics (95th edition), the enthalpies of combustion are as follows: formaldehyde, −566 kJ mol⁻¹; ribose, −2,349 kJ mol⁻¹; glucose, −2,840 kJ mol⁻¹.

The value of EE_{Formose} can be calculated as follows

$$EE_{\text{Formose}} = \left(-2,349 \text{ kJ mol}^{-1} \times \frac{1.6\% \text{ yield}}{5} - 2,840 \text{ kJ mol}^{-1} \times \frac{18.8\% \text{ yield}}{6} \right) / -566 \text{ kJ mol}^{-1} = 17.1\%$$

Thus, the global energy efficiency for the C₅₊ carbohydrates produced from the two-step electrochemical reduction–formose method can be estimated as

$$EE_{\text{global}} = EE_{\text{electrochemical}} \times EE_{\text{Formose}} = 4.8\% \times 17.1\% = 0.82\%$$

Data availability

All experimental data supporting the findings of this study are available in the Supplementary Information. Source data are provided with this paper.

References

- Crandall, B. S. et al. Transforming CO₂ into advanced 3D printed carbon nanocomposites. *Nat. Commun.* **15**, 10568 (2024).
- Belsa, B. et al. Materials challenges on the path to gigatonne CO₂ electrolysis. *Nat. Rev. Mater.* **9**, 535–549 (2024).
- Sullivan, I. et al. Coupling electrochemical CO₂ conversion with CO₂ capture. *Nat. Catal.* **4**, 952–958 (2022).
- Chen, Y. et al. Efficient multicarbon formation in acidic CO₂ reduction via tandem electrocatalysis. *Nat. Nanotechnol.* **19**, 311–318 (2024).
- Barecka, M. H., Ds Dameni, P., Zakir Muhamad, M., Ager, J. W. & Lapkin, A. A. energy-efficient ethanol concentration method for scalable CO₂ electrolysis. *ACS Energy Lett.* **8**, 3214–3220 (2023).
- Wu, Y., Liang, Y. & Wang, H. Heterogeneous molecular catalysts of metal phthalocyanines for electrochemical CO₂ reduction reactions. *Acc. Chem. Res.* **54**, 3149–3159 (2021).
- Ulmer, U. et al. Fundamentals and applications of photocatalytic CO₂ methanation. *Nat. Commun.* **10**, 3169 (2019).
- Ma, H. et al. Direct electroreduction of carbonate to formate. *J. Am. Chem. Soc.* **145**, 24707–24716 (2023).
- Zhang, X. et al. Molecular engineering of dispersed nickel phthalocyanines on carbon nanotubes for selective CO₂ reduction. *Nat. Energy* **5**, 684–692 (2020).
- Zhan, C. et al. Key intermediates and Cu active sites for CO₂ electroreduction to ethylene and ethanol. *Nat. Energy* **9**, 1485–1496 (2024).
- Soland, N. E., Roh, I., Huynh, W.-S. & Yang, P. Synthesis of carbohydrates from methanol using electrochemical partial oxidation over palladium with the integrated formose reaction. *ACS Sustain. Chem. Eng.* **11**, 12478–12483 (2023).
- Ruiz-Mirazo, K., Briones, C. & de la Escosura, A. Prebiotic systems chemistry: new perspectives for the origins of life. *Chem. Rev.* **114**, 285–366 (2014).
- Hann, E. C. et al. A hybrid inorganic–biological artificial photosynthesis system for energy-efficient food production. *Nat. Food* **3**, 461–471 (2022).
- Liu, Z., Wang, K., Chen, Y., Tan, T. & Nielsen, J. Third-generation biorefineries as the means to produce fuels and chemicals from CO₂. *Nat. Catal.* **3**, 274–288 (2020).
- Cestellos-Blanco, S. et al. Toward abiotic sugar synthesis from CO₂ electrolysis. *Joule* **6**, 2304–2323 (2022).

16. O'Brien, C. P., Watson, M. J. & Dowling, A. W. Challenges and opportunities in converting CO₂ to carbohydrates. *ACS Energy Lett.* **7**, 3509–3523 (2022).
17. Liu, G. et al. Solar-driven sugar production directly from CO₂ via a customizable electrocatalytic–biocatalytic flow system. *Nat. Commun.* **15**, 2636 (2024).
18. Zheng, T. et al. Upcycling CO₂ into energy-rich long-chain compounds via electrochemical and metabolic engineering. *Nat. Catal.* **5**, 388–396 (2022).
19. Cai, T. et al. Cell-free chemoenzymatic starch synthesis from carbon dioxide. *Science* **373**, 1523–1527 (2021).
20. Robinson, W. E., Daines, E., van Duppen, P., de Jong, T. & Huck, W. T. S. Environmental conditions drive self-organization of reaction pathways in a prebiotic reaction network. *Nat. Chem.* **14**, 623–631 (2022).
21. Bris, A. et al. Direct Analysis of complex reaction mixtures: formose reaction. *Angew. Chem. Int. Ed. Engl.* **63**, e202316621 (2024).
22. Haynes, W. M. et al. (eds) *CRC Handbook of Chemistry and Physics* 95th edn (CRC, 2014).
23. Li, J. et al. Mechanism-guided realization of selective carbon monoxide electroreduction to methanol. *Nat. Synth.* **2**, 1194–1201 (2023).
24. Wu, Y., Jiang, Z., Lu, X., Liang, Y. & Wang, H. Domino electroreduction of CO₂ to methanol on a molecular catalyst. *Nature* **575**, 639–642 (2019).
25. Wu, Y., Jiang, Z., Lin, Z., Liang, Y. & Wang, H. Direct electrosynthesis of methylamine from carbon dioxide and nitrate. *Nat. Sustain.* **4**, 725–730 (2021).
26. Singh, A. et al. Molecular electrochemical catalysis of CO-to-formaldehyde conversion with a cobalt complex. *J. Am. Chem. Soc.* **146**, 22129–22133 (2024).
27. Chan, T. et al. Role of mass transport in electrochemical CO₂ reduction to methanol using immobilized cobalt phthalocyanine. *ACS Appl. Energy Mater.* **7**, 3091–3098 (2024).
28. Zhu, Q. S. et al. The solvation environment of molecularly dispersed cobalt phthalocyanine determines methanol selectivity during electrocatalytic CO reduction. *Nat. Catal.* **7**, 987–999 (2024).
29. Cheon, S., Li, J. & Wang, H. In situ generated CO enables high-current CO₂ reduction to methanol in a molecular catalyst layer. *J. Am. Chem. Soc.* **146**, 16348–16354 (2024).
30. Li, J., Al-Mahayni, H., Chartrand, D., Seifitokaldani, A. & Kornienko, N. Electrochemical formation of C–S bonds from CO₂ and small-molecule sulfur species. *Nat. Synth.* **2**, 757–765 (2023).
31. Zhang, X. et al. Highly selective and active CO₂ reduction electrocatalysts based on cobalt phthalocyanine/carbon nanotube hybrid structures. *Nat. Commun.* **8**, 14675 (2017).
32. Kim, H. J. et al. Synthesis of carbohydrates in mineral-guided prebiotic cycles. *J. Am. Chem. Soc.* **133**, 9457–9468 (2011).
33. Delidovich, I. V., Simonov, A. N., Taran, O. P. & Parmon, V. N. Catalytic formation of monosaccharides: from the formose reaction towards selective synthesis. *ChemSusChem* **7**, 1833–1846 (2014).
34. Iqbal, Z. & Novalín, S. The formose reaction: a tool to produce synthetic carbohydrates within a regenerative life support system. *Curr. Org. Chem.* **16**, 769–788 (2012).
35. Omran, A., Menor-Salvan, C., Springsteen, G. & Pasek, M. The messy alkaline formose reaction and its link to metabolism. *Life* **10**, 125 (2020).
36. Nielsen, S. S. in *Food Analysis Laboratory Manual, Food Science Texts Series* (ed. Nielsen, S. S.) 47–53 (Springer US, 2010).
37. Iqbal, M. Z. & Novalín, S. Analysis of formose sugar and formaldehyde by high-performance liquid chromatography. *J. Chromatogr. A* **1216**, 5116–5121 (2009).
38. Angyal, S. J. The Lobry de Bruyn-Alberda van Ekenstein transformation and related reactions. *Top. Curr. Chem.* **215**, 1–14 (2001).
39. Waki, M., Shirai, S. & Hase, Y. Saccharide formation by sustainable formose reaction using heterogeneous zeolite catalysts. *Dalton Trans.* **53**, 2678–2686 (2024).
40. Yue, H. R., Zhao, Y. J., Ma, X. B. & Gong, J. L. Ethylene glycol: properties, synthesis, and applications. *Chem. Soc. Rev.* **41**, 4218–4244 (2012).
41. Weinberg, N. L. & Mazur, D. J. Electrochemical hydrodimerization of formaldehyde to ethylene-glycol. *J. Appl. Electrochem.* **21**, 895–901 (1991).
42. Xia, R. et al. Electrosynthesis of ethylene glycol from C₁ feedstocks in a flow electrolyzer. *Nat. Commun.* **14**, 4570 (2023).
43. Li, R., Yang, D. J., Lou, H. M., Zhou, M. S. & Qiu, X. Q. Influence of sulfonated acetone-formaldehyde condensation used as dispersant on low rank coal-water slurry. *Energy Convers. Manage.* **64**, 139–144 (2012).
44. Li, Y. J. et al. The molecular structure and spectroscopic properties of formaldoxime (CH₂NOH). *Phys. Scr.* **99**, 055403 (2024).

Acknowledgements

This work was supported by the National Brown Investigator Award (H.W.) and US National Science Foundation (grant number CHE-2154724, H.W.). The flow cell work was supported by the Yale Center for Natural Carbon Capture (H.W.).

Author contributions

J.L. and H.W. conceived this project and designed the experiments. J.L. and K.C. synthesized the catalyst materials, performed the TEA and LCA and conducted the electrochemical and chemical reactions. Y.S. assisted in some of these experiments. J.Y. carried out the post-reaction separation of carbohydrates. J.L., K.C. and H.W. wrote the manuscript with input from N.E.S. and P.Y. The manuscript was edited by Y.G. and S.C., who also contributed to data analysis. H.W. supervised the project.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s44160-025-00961-x>.

Correspondence and requests for materials should be addressed to Peidong Yang or Hailiang Wang.

Peer review information *Nature Synthesis* thanks Rong Xia and the other, anonymous, reviewer(s) for their contribution to the peer review of this work. Primary Handling Editor: Alexandra Groves, in collaboration with the *Nature Synthesis* team.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

© The Author(s), under exclusive licence to Springer Nature Limited 2026